Two separate and distinct syntheses of stereospecifically deuteriated samples of (2S)-proline^{\dagger}

Paul Barraclough, Petra Dieterich, Caroline A. Spray and Douglas W. Young*

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Two distinct syntheses of samples of the amino acid L-proline which are stereospecifically deuteriated on the β -carbon atom are reported. In the first of these, the labelled diazoketones **6**, prepared by a chemico-enzymatic synthesis, have been photolysed in alkaline conditions to give the corresponding labelled methyl pyroglutamates **10** via hydrolysis and intramolecular trapping of the resultant ketene intermediates **9**. These were then converted into (2S,3S)- $[3-^2H_1]$ - and (2S,3R)- $[2,3-^2H_2]$ -proline, **1a** and **1b** respectively. The second synthesis provides (2S)- $[3,3-^2H_2]$ -, (2S,3S)- and (2S,3R)- $[3-^2H_1]$ -proline, **1d**, **1a** and **1c** respectively, and has as its key step the highly stereoselective hydrolysis of the silylenol ethers **14** and **14a** respectively in which deuteriation or protonation occurs from the *re*-face of the enol ether.

Introduction

The use of stereospecifically labelled amino acids in metabolic studies² and, in combination with multidimensional NMR spectroscopy, in obtaining detailed protein solution structures³⁻⁵ makes the ready availability of such compounds of considerable importance. The amino acid L-proline 1 and its post-translationally modified derivatives are constituents of antibiotics and are important for conformational constraint in proteins. Several syntheses of stereospecifically labelled samples of this amino acid have been completed by ourselves¹ and others^{2,6} and we now wish to report two straightforward syntheses of samples of L-proline which are labelled with deuterium in the 3-*pro-R* or the 3-*pro-S* position.



Department of Chemistry, University of Sussex, Falmer, Brighton, UK BN1 9QJ † See ref. 1.

Results and discussion

As the first step in our synthesis of stereospecifically labelled Lamino acids⁷ and β -amino acids,⁸ we have used the well-known^{2,9} anti-addition of ammonia to the double bond of fumaric acid 2 by the enzyme L-aspartase (EC 4.3.1.1) to obtain samples of (2S, 3R)- $[3-^{2}H_{1}]$ - and $(2S,3S)-[2,3-^{2}H_{2}]$ -aspartic acid, **3a** and **3b** respectively, as shown in Scheme 1. We subsequently found that using this enzyme in a preparation of Escherichia coli immobilised on polyacrylamide gel¹⁰ was more effective for accessing these labelled samples 3 of aspartic acid which were used in our general synthesis of stereospecifically labelled amino acids.11 In our synthesis of labelled glutamic acids,^{7a,b} we treated the labelled aspartic acids **3** with trifluoroacetic anhydride, followed by methanol to obtain a mixture containing *ca*. 80% of the trifluoroacetyl- β -acid- α -ester 4 together with *ca*. 20% of its isomer, the α -ester- β -acid. The mixed esters were converted into a mixture of the acid chloride 5 and its isomer which were originally^{7b} separated by recrystallisation from benzene. The isomer 5 was then converted into the diazoketone 6. In the current synthesis, we have carried through the mixtures of regioisomers to the diazoketones 6, 6a, and 6b which have been



Scheme 1 Reagents and conditions: (i) see ref. 7b; (ii) hv, MeOH, 3 h (96%); (iii) hv, MeOH, aq. NaHCO₃, 3 h (quantitative).

purified by recrystallisation from dichloromethane and petroleum ether.

In our synthesis of labelled samples of glutamic acid,^{7a,b} the diazoketones 6 were photolysed in redistilled methanol to give labelled samples of dimethyl N-trifluoroacetylglutamate 8 via rearrangement to the ketenes 7 which reacted with the solvent methanol. However, when we carried out the photolysis of the unlabelled diazoketone 6 in methanol which had previously been dried by distillation from magnesium methoxide or calcium hydride, a new product was obtained. This was identified as methyl pyroglutamate 10 by spectroscopic comparison with an authentic sample. When the reaction was repeated using the labelled diazoketones, 6a and 6b, stereospecifically labelled samples of methyl pyroglutamate 10a and 10b were obtained. The most reasonable explanation for the new course of the reaction was that the solution had become slightly alkaline, leading to hydrolysis of the trifluoroacetate to give the primary amine 9. The amine would then undergo intramolecular nucleophilic addition to the ketene function, yielding the pyroglutamate 10. In neutral conditions the unhydrolysed trifluoroacetamide moiety would not be sufficiently nucleophilic to compete with methanol for the ketene in the intermediate 7 and so the protected dimethyl glutamate 8 would be formed. To test this hypothesis, the diazoketone 6 was hydrolysed to the corresponding free amino acid by adaptation of a literature method.¹² Photolysis of the crude amino acid diazoketone in methanol gave (2S)-pyroglutamic acid. However, when this sequence of reactions was conducted using the stereospecifically labelled samples of diazoketone, 6a and 6b, the 3R and 3Sdeuterium labels (but not the C-2 label) were exchanged. Photolysis in methanol containing sodium methoxide caused exchange of labels both at C-2 and at C-3. Eventually, conditions were found which reliably and consistently gave the stereospecifically labelled methyl pyroglutamates 10a and 10b in quantitative yield. This involved photolysis in methanol containing 1 equivalent of 6% w/v aqueous sodium bicarbonate.

The samples of methyl pyroglutamate, **10a** and **10b**, obtained by this latter method, were converted into the corresponding *tert*butoxycarbonyl protected derivatives **11a** and **11b** by reaction with di-*tert*-butyl dicarbonate and DMAP in acetonitrile, as shown in Scheme 2. Reduction using borane dimethylsulfide then gave the protected proline derivatives **12a** and **12b**. The ¹H NMR spectra of the products **12** were complicated by the well-known¹³ conformational isomerism found in *N*-acylproline derivatives but, when the products were deprotected using refluxing 6 M aqueous HCl, good yields of the labelled prolines **1a** and **1b** were obtained. The ¹H NMR spectra of these compounds, shown in Fig. 1, indicated stereospecific labelling, and so synthesis of (2S,3S)- $[3-^2H_1]$ -proline **1a** and (2S,3R)- $[2,3-^2H_2]$ -proline **1b** had been achieved.

In our second synthesis, where stereoselectively β -monodeuteriated products were required, we used the silylenol ether **14**, prepared during our work on the synthesis of analogues of kainic acid¹⁴ as starting material. When this silylenol ether was prepared from a solution of the ketone **13**¹⁴ using excess trimethylsilyl chloride and the product was treated *in situ* with ²H₂O (Scheme 3), we observed remarkable stereoselectivity in deuteriation at C-3 in the resultant ketone **13a**, since, although the ¹H NMR spectrum of the unpurified product was complicated by the conformational isomerism¹³ shown by *N*-acylprolines,



Scheme 2 Reagents and conditions: (i) $O(CO_2'Bu)_2$, DMAP, CH₃CN, rt, overnight (71% 11, 24% 11a, 26% 11b); (ii) H₃B·SMe₂, THF, 0 °C, 18.5 h (65% 12, 66% 12a, 56% 12b); (iii) 6 M HCl, reflux, 2 h (55% 1, 78% 1a, 94% 1b).



Fig. 1 360 MHz ¹H NMR spectra in $C^2H_3O^2H$ of (a) (2*S*)-proline **1**; (b) (2*S*,3*S*)-[3-²H₁]-proline **1a**; and (c) (2*S*,3*R*)-[2,3-²H₂]-proline **1b**.

only one of the two multiplets at δ 2.50 and 2.89 ppm (due to the diastereotopic protons at C-3) was absent in the ¹H NMR spectrum of the ketone **13a**. Since the signal at δ 2.89 ppm showed a 4.5% enhancement upon irradiation of the signal for H-2 at *ca*. δ 4.6 ppm, it was evidently due to H-3*R* and the signal at δ 2.50 ppm was therefore assigned as being due to H-3*S*. Deuteriation had evidently occurred from the *re*-face of the silylenol ether **14**. Attempts to purify the compound by chromatography on silica



Scheme 3 Reagents and conditions: (i) (a) LDA, THF, -78 °C; (b) Me₃SiCl, -78 °C; (ii) ²H₂O, rt (quantitative); (iii) NaBH₄, MeOH, Et₂O, 0 °C, 5 min, then rt, 30 min (98% 15 from 13, 85% 15a from 13a).

gel 'washed out' the label. The unpurified compounds 13 and 13a were reduced directly to the stable alcohols 15 and 15a respectively using sodium borohydride in methanol and diethyl ether at 0 °C, as shown in Scheme 3. This allowed the stereoselectivity to be confirmed using a purer and more stable compound. The ¹H NMR spectrum of the purified unlabelled compound 15 in C²HCl₃ was complicated by conformational isomerism but a simpler spectrum was obtained for a solution in C²H₃CN at 60 °C. NOE experiments in C²HCl₃, summarised in Fig. 2, allowed assignment of the signals due to the hydrogens H-3R and H-3S, since irradiation at δ 4.2 ppm (H-2) gave a considerably larger enhancement to the signal for H-3 at δ 2.3 ppm than to that at δ 2.0 ppm. The former signal was therefore assigned as being due to H-3R. Irradiation at the signal at δ 4.3 ppm (H-4) gave a large enhancement to the signal for H-3*R*, thus confirming the stereochemistry at C-4. The 1 H NMR spectrum of the labelled alcohol 15a exhibited a broad one-proton singlet for H-3S at δ 2.03 ppm and a residual multiplet (*ca.* 20%) for H-3R at δ 2.3 ppm. The stereoselectivity of hydrolysis of the silvlenol ether 14 was thus confirmed. No deuterium was present in H-3S and the small amount of protium at H-3R was presumably the result of an isotope effect.



Fig. 2 NOE experiments on the alcohol 15.

The high stereoselectivity found in this reaction was somewhat surprising and it is tempting to suggest that the mechanism shown in Scheme 4 could be partly synchronous giving the chair-like transition state **16B** shown. Interaction of the bulky trimethylsilyl and *tert*-butoxycarbonyl groups in **16B** would result in deuteriation occurring from the *re*-face as shown.

Reasoning that we might obtain the isotopomer 13b by protonation of the deuteriated silvlenol ether 13a, we decided to prepare the $[3,3-{}^{2}H_{2}]$ -ketone 13c. We therefore treated the ketone 13 with lithium diisopropylamide (LDA) in tetrahydrofuran and quenched the solution with ²H₂O. The ¹H NMR spectrum of the product indicated that any label introduced at C-3 had been 'washed out' upon purification using silica gel chromatography but that there was evidently considerable deuteriation (ca. 75%) at C-5. We therefore stirred the ketone 13 in ${}^{2}H_{2}O$ containing silica gel which had been washed with ²H₂O (Scheme 5). After 10 d at room temperature, complete exchange was obtained and the product 13c was converted to a solution of the silylenol ether 14a by reaction with LDA followed by treatment with excess trimethylsilyl chloride. Addition of H₂O to the solution gave the ketone 13b. The ¹H NMR spectrum of this compound exhibited a signal at δ 2.89 ppm for H-3*R* but not at δ 2.50 ppm for H-3*S*.

Since the ketone **13** had been shown to be so acidic, readily exchanging at C-3, the labelled sample **13b** was not further purified, but reduced immediately with sodium borohydride to give the alcohol **15b**. The ¹H NMR spectrum of the purified alcohol **15b** showed a one-proton multiplet at δ 2.28 ppm for H-3*R* and *ca*. 30% residual protium at δ 2.0 ppm for H-3*S*. The residual protium presumably reflected incomplete deuteriation in the initial dideuteriated compound.



Scheme 5 Reagents and conditions: (i) $^{2}H_{2}O$, silica gel, rt, 10 d (93%); (ii) (a) LDA, THF, $-78 \,^{\circ}C$, (b) Me₃SiCl, $-78 \,^{\circ}C$; (iii) H₂O, rt (quantitative); (iv) NaBH₄, MeOH, Et₂O, 0 $^{\circ}C$, 5 min, then rt, 30 min (40% 15b from 13b; 89% 15c from 13c).

Conversion of the unlabelled and labelled alcohols **15** to the corresponding samples of proline **1** proceeded as described in Scheme 6. The alcohols **15** were first converted into the *para*-toluenesulfonates **17** using *para*-toluenesulfonyl chloride in pyridine at room temperature. These were then reduced to the protected proline derivatives **18** by heating at 85 °C with NaBH₄ in DMSO for 6.5 h. Deprotection using 6 M aqueous hydrochloric acid for 2 h at room temperature gave the samples of proline, **1**, **1a**, **1c** and **1d**. The ¹H NMR spectra of these compounds are shown in Fig. 3, the sample of (2S,3S)- $[3-^2H_1]$ -proline **1a** having 22% protium at H-3*S*, and that of (2S,3R)- $[3-^2H_1]$ -proline **1c** having *ca*. 33% protium at H-3*R*. The same synthetic route was used to convert (2S)- $[3,3-^2H_2]$ -4-ketoproline **15c** to (2S)- $[3,3-^2H_2]$ proline **1d**.



Scheme 6 *Reagents and conditions:* (i) TsCl, pyridine, rt, 20 h (58% 17, 33% 17a, 71% 17b, 50% 17c); (ii) NaBH₄, DMSO, 85 °C, 6.5 h (94% 18, 87% 18a, 59% 18b, 89% 18c); (iii) 6 M HCl, rt, 2 h (88% 1, 27% 1a, 71% 1c, 53% 1d).

Conclusions

Two distinct and separate syntheses of samples of L-proline which are stereospecifically deuteriated in the β -position have been completed. The first, a chemico-enzymatic synthesis, provided samples of (2S,3S)- $[3-^2H_1]$ - and (2S,3R)- $[2,3-^2H_2]$ -proline, **1a** and **1b**, while the second, a chemical synthesis from a homochiral 4-ketoproline derivative, provided samples of (2S,3S)- and (2S,3R)- $[3-^2H_1]$ -proline, **1a** and **1c**.

Experimental

Melting points were determined on a Kofler hot-stage and are uncorrected. Optical rotations (given in units of $10^{-1} \text{ deg cm}^{-2} \text{ g}^{-1}$)



Fig. 3 360 MHz ¹H NMR spectra in $C^2H_3O^2H$ of (a) (2*S*)-proline 1; (b) (2*S*,3*S*)-[3-²H₁]-proline 1a; (c) (2*S*,3*R*)-[3-²H₁]-proline 1c; and (d) (2*S*)-[3,3-²H₂]-proline 1d.

were measured on a Perkin Elmer PE241 polarimeter using a 1dm path length micro cell. IR spectra were recorded on a Perkin Elmer 1710 Fourier transform instrument. ¹H NMR spectra were recorded on Bruker WM360 (360 MHz), AMX500 (500 MHz) and AC-P250 (250 MHz) instruments. ¹³C NMR spectra were recorded on Bruker AMX 500 (125.8 MHz) WM360 (90.6 MHz) and AC-P250 (62.9 MHz) instruments. J values are given in Hz. Residual solvent peaks were used as internal references in all NMR spectra. Low resolution mass spectra were recorded at Sussex University using Kratos MS80RF and Fisons VG Autospec instruments by Mr A. M. Greenway and Dr A. Abdul Sada, and at the Wellcome Research Laboratories, Beckenham using Kratos Concept 1S and MS50, and Fisons BIO Q instruments by Dr S. Chotai. High resolution mass measurements were performed by Dr S. Chotai at the Wellcome Research Laboratories, Beckenham using a Kratos Concept 1S instrument. Microanalyses were performed by Miss M. Patel at Sussex University, and by Ms C. Lawless and W. C. Man at the Wellcome Research Laboratories, Beckenham. Column chromatography was carried out using Merck Kieselgel 60 (230-400 mesh-Art 9385). Unless otherwise indicated, petroleum ether refers to that fraction of hexanes of bp 60-80 °C.

Dimethyl (2S)-N-trifluoroacetylglutamate (8)

Methyl (2*S*)-5-diazo-4-oxo-trifluoroacetamidopentanoate 6^{7b} (500 mg, 1.87 mmol) was dissolved in methanol (200 ml) that had been purified by fractional distillation. The solution was degassed under a rapid stream of dry nitrogen for 30 min and then irradiated using a 125 W medium pressure Hanovia UV lamp fitted with a quartz filter. The reaction was shown to be complete after 3 h by UV spectroscopy. The solvent was removed *in vacuo* to give dimethyl (2*S*)-*N*-trifluoroacetylglutamate **8** (460 mg, 96%) as a brown oil with identical spectra to those of an authentic sample.

Methyl (2S)-pyroglutamate (10)

Methyl (2*S*)-5-diazo-4-oxo-2-trifluoroacetamidopentanoate 6^{7b} (3 g, 11.2 mmol) was added to a mixture of methanol (100 ml) and aqueous sodium hydrogencarbonate (6% w/v, 1.56 ml). Further

methanol (500 ml) was added and the solution was degassed using a rapid stream of dry argon with continuous stirring for 45 min. Precipitated bicarbonate had dissolved after this time and the mixture was irradiated using a 125 W Hanovia medium pressure UV lamp fitted with a quartz filter until no starting material could be detected by UV spectroscopy (ca. 3 h). The solvent was removed in vacuo to give methyl (2S)-pyroglutamate 10 as a viscous yellow oil after thorough drying (1.6 g, quant.). A sample was purified by column chromatography on silica gel by sequential elution with ethyl acetate-petroleum ether (1:2); then ethyl acetate; then ethyl acetate-methanol (9:1) to yield methyl (2S)-pyroglutamate 10 as a pale yellow oil; $[a]_{D}^{22} - 2.3 (c 1, CH_2Cl_2)$ [lit.¹⁵ - 6.95 (c 1, CH_2Cl_2)]; $v_{\rm max}$ (film)/cm⁻¹ 3300 (br, NH), 1742 (ester) and 1698 (lactam); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 2.18–2.39 (4H, m, H-3 and H-4), 3.76 (3H, s, OCH₃), 4.26 (1H, dd, *J*_{2.3*R*} 5, *J*_{2.3*S*} 9, H-2) and 6.40 (1H, br s, NH, exch. in ${}^{2}H_{2}O$); δ_{C} (90.6 MHz, $C^{2}H_{3}O^{2}H$) 26.2 (C-3), 30.6 (C-4), 53.4 (OCH₃), 57.3 (C-2), 174.9 (ester) and 181.0 (lactam).

Methyl (2*S*,3*S*)-[3-²H₁]-pyroglutamate (10a)

This was prepared from methyl (2*S*,3*R*)-[3-²H₁]-5-diazo-4-oxo-2trifluoroacetamidopentanoate **6a** ^{7b} (3.1 g, 11.6 mmol) as described above (1.67 g, quant.). A sample was purified by chromatography as described above; v_{max} (film)/cm⁻¹ 3380 (br, NH), 1730 (br, ester) and 1686 (lactam); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 2.18–2.37 (3H, m, H-3*R* and H-4), 3.75 (3H, s, OCH₃), 4.23 (1H, d, $J_{2,3R}$ 5, H-2) and 6.60 (1H, br s, NH); $\delta_{\rm C}$ (62.9 MHz, C²H₃O²H) 26.0 (t, C-3), 30.6 (C-4), 53.4 (OCH₃), 57.4 (C-2), 174.9 (ester) and 181.5 (lactam).

Methyl (2S,3R)- $[2,3-^{2}H_{2}]$ -pyroglutamate (10b)

This was prepared from methyl (2*S*,3*S*)-[2,3⁻²H₂]-5-diazo-4-oxo-2trifluoroacetamidopentanoate **6b**^{7b} (3.6 g, 13.4 mmol) as described above (1.88 g, 97%). A sample was purified by chromatography as described above; v_{max} (film)/cm⁻¹ 3380 (br, NH), 1730 (ester) and 1681 (lactam); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 2.30–2.50 (3H, m, H-3 and H-4), 3.77 (3H, s, OCH₃) and 6.1 (1H, br s, NH); $\delta_{\rm C}$ (90.6 MHz, C²H₃O²H) 25.8 (t, C-3), 30.6 (C-4), 53.3 (OCH₃), 57.0 (t, C-2), 174.9 (ester) and 181.4 (lactam).

Methyl (2S)-N-tert-butoxycarbonylpyroglutamate (11)

Crude methyl (2S)-pyroglutamate 10 from photolysis of the diazoketone 6 (1.6 g, 11.2 mmol) was dissolved in acetonitrile (50 ml) at 0 °C under nitrogen. DMAP (122 mg, 1 mmol) was added and the solution was stirred at room temperature for 10 min. Ditert-butyl dicarbonate (3.17 g, 14.5 mmol) was added and stirring was continued overnight at room temperature. The solvent was removed in vacuo to afford an orange gum which was dissolved in ethyl acetate (50 ml). The organic layer was washed with saturated aqueous sodium hydrogen carbonate (2×20 ml), 10% aqueous citric acid $(2 \times 20 \text{ ml})$ and saturated aqueous sodium chloride (20 ml), and dried (MgSO₄). The solvent was removed *in vacuo* to give an orange solid which was purified by column chromatography on silica gel, eluting with diethyl ether to yield methyl (2S)-N-tertbutoxycarbonylpyroglutamate **11** as a white solid (2.73 g, 71%); mp 69 °C (lit.¹⁶ 72.5–73.5 °C); [a]²² –30 (c 0.36, CHCl₃) [lit.¹⁶ -30.4 (c 1.0, CHCl₃)]; (Found C, 54.3; H, 7.0; N, 5.7. C₁₁H₁₇NO₅ requires C, 54.3; H, 7.0; N, 5.8%); m/z [+ve FAB, (CHCl₃/NBA)], 244 ($[M + H]^+$); v_{max} (KBr)/cm⁻¹ 1760 (imide), 1741 (ester) and 1705 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.42 [9H, s, C(CH₃)₃], 1.90 (1H, ddd, $J_{3R,3S}$ 13.3, $J_{3R,2}$ 3.0, $J_{3R,4R}$ 10.0, $J_{3R,4S}$ 3.7, H-3*R*), 2.30 (1H, m, $J_{3S,3R}$ 13.3, $J_{3S,2} \approx J_{3S,4R} \approx J_{3S,4S} \approx 9.4$, H-3*S*), 2.40 (1H, ddd, $J_{4S,4R}$ 17.5, $J_{4S,3S}$ 9.4, $J_{4S,3R}$ 3.7, H-4*S*), 2.60 (1H, m, $J_{4R,4S}$ 17.5 $J_{4R,3S} \approx J_{4R,3R} \approx 10.0$, H-4*R*), 3.72 (3H, s, OCH₃) and 4.55 (1H, dd, $J_{2,3S}$ 9.4, $J_{2,3R}$ 3.0, H-2); irradiation at δ 4.55 ppm (H-2) gave 3.5% enhancement at δ 2.30 ppm (H-3*S*) and 1% enhancement at δ 1.90 ppm (H-3*R*); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 21.5 (C-3), 27.9 [C(CH₃)₃], 31.1 (C-4), 52.5 (OCH₃), 58.8 (C-2), 83.6 [OC(CH₃)₃], 149.3 (urethane), 171.8 (ester) and 172.9 (lactam).

Methyl (2*S*,3*S*)-[3-²H₁]-*N*-tert-butoxycarbonylpyroglutamate (11a)

This was prepared by as described above from methyl (2*S*,3*S*)-[3-²H₁]-pyroglutamate **10a** (4.3 g, 29.8 mmol) (1.73 g, 24%); mp 68– 70 °C; $[a]_{D}^{22}$ –31.2 (*c* 0.35, CHCl₃); *m*/*z* [+ve FAB, (CHCl₃/NBA)], 245 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 1759 (imide), 1742 (ester) and 1704 (urethane); δ_{H} (360 MHz, C²HCl₃) 1.49 [9H, s, C(CH₃)₃], 2.0 (1H, br d, $J_{3R,4R}$ 9.6, H-3*R*), 2.50 (1H, dd, $J_{4S,4R}$ 17.5, $J_{4S,3R}$ 3.6, H-4*S*), 2.60 (1H, dd, $J_{4R,4S}$ 17.5, $J_{4R,3R}$ 9.6, H-4*R*), 3.79 (3H, s, OCH₃) and 4.60 (1H, d, $J_{2,3R}$ 2.8, H-2); δ_{C} (125.8 MHz, C²HCl₃) 21.1 (t, C-3), 27.8 [C(CH₃)₃], 30.9 (C-4), 52.4 (OCH₃), 58.7 (C-2), 83.5 [O*C*(CH₃)₃], 149.2 (urethane), 171.8 (ester) and 172.9 (lactam).

Methyl (2*S*,3*R*)-[2,3-²H₂]-*N*-tert-butoxycarbonylpyroglutamate (11b)

This was prepared as described above from methyl (2*S*,3*R*)-[2,3⁻²H₂]-pyroglutamate **10b** (5.59 g, 38.5 mmol) as a solid (2.45 g, 26%); mp 69–70 °C; $[a]_{D}^{22}$ –31.5 (*c* 0.42, CHCl₃); *m/z* [+ve FAB, (CHCl₃/NBA)], 246 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 1759 (imide), 1740 (ester) and 1704 (urethane); δ_{H} (360 MHz, C²HCl₃) 1.47 [9H, s, C(CH₃)₃] 2.20 (1H, br t, *J* 10, H-3*S*), 2.50 (1H, dd, *J*_{4*S*,4*R*} 17.5, *J*_{4*S*,35} 9.4, H-4*S*), 2.60 (1H, dd, *J*_{4*R*,45} 17.5, *J*_{4*R*,35} 10.2, H-4*R*) and 3.77 (3H, s, OCH₃); δ_{C} (125.8 MHz, C²HCl₃) 21.1 (t, C-3), 27.8 [C(CH₃)₃], 30.9 (C-4), 52.3 (OCH₃), 58.4 (t, C-2), 83.4 [OC(CH₃)₃], 149.2 (urethane), 171.7 (ester) and 172.9 (lactam).

Methyl (2S)-N-tert-butoxycarbonylprolinate (12)

Methyl (2S)-N-tert-butoxycarbonylpyroglutamate 11 (180 mg, 0.74 mmol) was dissolved in dry tetrahydrofuran (10 ml) and the solution was cooled to 0 °C. Borane dimethylsulfide (2 M in tetrahydrofuran, 0.7 ml, 1.4 mmol) was added via syringe and the mixture was stirred for 18.5 h. The solvent was removed in vacuo and the crude oily product was dissolved in ethyl acetate (20 ml). The organic layer was washed with water (2 \times 10 ml) and saturated aqueous sodium chloride (10 ml), dried (MgSO₄), filtered and the solvent was removed in vacuo to yield methyl (2S)-*N-tert*-butoxycarbonylprolinate **12** as a clear oil (110 mg, 65%); $[a]_{D}^{22}$ -61.7 (c 1.15, MeOH) [lit.¹⁷ -68.3 (c 1.08, MeOH) for the [4,4- 2 H₂]-labelled compound]; m/z [FAB/3-NBA] 252 ([M + Na]⁺) and 230 ($[M + H]^+$); m/z [EI], 170 ($[M - CO_2CH_3]^+$); v_{max} (film)/cm⁻¹ 1751 (ester) and 1703 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.36 and 1.42 [9H, 2s, C(CH₃)₃], 1.80 (3H, m, H-3R and H-4), 2.20 (1H, m, H-3S), 3.40 (2H, m, H-5), 3.70 (3H, s, OCH₃), and 4.18 and 4.28 $(1H, 2dd, H-2); \delta_{C} (125.8 \text{ MHz}, C^{2}HCl_{3}) 23.6 \text{ and } 24.2 (C-4), 28.2$ and 28.3 [C(CH₃)₃], 29.8 and 30.8 (C-3), 46.2 and 46.5 (C-5), 52.0 (OCH_3) , 58.6 and 59.0 (C-2), 79.7 and 79.9 $[OC(CH_3)_3]$, 153.7 and 154.3 (urethane), and 173.4 and 173.7 (ester).

Methyl (2S,3S)-[3-²H₁]-N-tert-butoxycarbonylprolinate (12a)

This was prepared as above from methyl (2S,3S)-[3- $^{2}H_{1}]$ -*Ntert*-butoxycarbonylpyroglutamate **11a** (74 mg, 0.30 mmol) and purified by column chromatography on silica gel, eluting with 1 : 1 ethyl acetate–petroleum ether (40–60 °C) (46 mg, 66%); $[a]_{D}^{23} - 58.2$ (*c* 0.98, MeOH); *m*/*z* [EI], 171 [M – CO₂CH₃]⁺; v_{max} (film)/cm⁻¹ 1751 (ester) and 1703 (urethane); δ_{H} (360 MHz, C²HCl₃) 1.37 and 1.43 [9H, 2s, C(CH₃)₃], 1.80 (3H, m, H-3*R* and H-4), 3.40 (2H, m, H-5), 3.69 (3H, s, OCH₃) and 4.28 and 4.31 (1H, 2br s, H-2); δ_{C} (125.8 MHz, C²HCl₃) 23.5 and 24.1 (C-4), 28.2 and 28.3 [C(*C*H₃)₃], 29.0 and 30.0 (t, C-3), 46.2 and 46.5 (C-5), 52.0 (OCH₃), 58.6 and 59.0 (C-2), 79.7 and 79.9 [OC(CH₃)₃], 153.7 and 154.4 (urethane) and 173.4 and 173.7 (ester).

Methyl (2S,3R)-[2,3-²H₂]-N-tert-butoxycarbonylprolinate (12b)

This was prepared as above from methyl (2S,3R)- $[2,3-^{2}H_{2}]$ -*N*-*tert*-butoxycarbonylpyroglutamate **11b** (80 mg, 0.326 mmol) and purified by column chromatography on silica gel, eluting with 1 : 1 ethyl acetate–petroleum ether (40–60 °C) (42 mg, 56%); $[a]_{D}^{25}$ – 56.5 (*c* 0.96, MeOH); *m/z* [EI], 172 ([M – CO₂CH₃]⁺); v_{max} (film)/cm⁻¹ 1751 (ester) and 1703 (urethane); δ_{H} (360 MHz, C²HCl₃) 1.39 and 1.44 [9H, 2s, C(CH₃)₃], 1.80 (2H, m, H-4), 2.20 (1H, m, H-3S), 3.40 (2H, m, H-5) and 3.70 (3H, s, OCH₃); δ_{C} (125.8 MHz, C²HCl₃) 23.5 and 24.2 (C-4), 28.2 and 28.4 [C(CH₃)₃], 29.1 and 30.4 (t, C-3), 46.2 and 46.5 (C-5), 52.0 (OCH₃), 58.6 (m, C-2), 79.7 and 79.8 [OC(CH₃)₃], 153.7 and 154.4 (urethane), and 173.5 and 173.7 (ester).

(2S)-Proline (1)

Methyl (2S)-N-tert-butoxycarbonylprolinate 12 (98 mg, 0.427 mmol) was heated at reflux with 6 M aqueous HCl (2 ml) for 2 h. The product was dried by lyophilisation and excess water was removed by vacuum desiccation. The residue was dissolved in water, the pH was adjusted from 2 to 7 with 2 M aqueous ammonium hydroxide, and the solution was loaded onto a Dowex 50X8 ion exchange column. The column was washed with water until the eluant was no longer acidic. The product 1 was eluted with 2 M aqueous ammonium hydroxide, dried by lyophillisation and obtained as an off-white solid (30 mg, 55%); mp 229 °C (decomp.); $(lit.^{15} 227-229 \ ^{\circ}C); [a]_{D}^{25} - 82.0 (c 0.72, H_2O) [lit.^{15} - 83 (c 1, H_2O)];$ m/z [+ve FAB (thioglycerol)], 231 ([2M + H]⁺), 138 ([M + Na]⁺) and 116 ($[M + H]^+$); v_{max} (KBr)/cm⁻¹ 3400 (br, NH and OH) and 1622 (br, carboxylate); $\delta_{\rm H}$ (360 MHz, C²H₃O²H) 1.96 (2H, m, H-4), 2.13 (1H, m, H-3R), 2.28 (1H, m, H-3S), 3.23 (1H, m, H-5A), 3.36 (1H, m, H-5B) and 3.95 (1H, dd, J_{2,3R} 6.2, J_{2,3S} 8.7, H-2); δ_c (62.9 MHz, C²H₃O²H) 25.1 (C-4), 30.4 (C-3), 47.0 (C-5), 62.6 (C-2) and 174.2 (acid).

(2*S*,3*S*)-[3-²H₁]-Proline (1a)

This was prepared as above from methyl (2S,3S)- $[3-^2H_1]$ -*N-tert*butoxycarbonylprolinate **12a** (46 mg, 0.20 mmol) (20 mg, 78%); mp 230 °C (decomp.); $[a]_D^{25}$ -83.2 (*c* 0.1, H₂O); *m/z* [+ve FAB (thioglycerol)], 117 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 3400 (br, NH and OH) and 1626 (carboxylate); $\delta_{\rm H}$ (360 MHz, C²H₃O²H) 1.95 (2H, m, H-4), 2.20 (1H, m, H-3*R*), 3.23 (1H, m, H-5A), 3.36 (1H, m, H-5B) and 3.94 (1H, d, $J_{2,3R}$ 6.2, H-2); $\delta_{\rm C}$ (62.9 MHz, C²H₃O²H) 25.4 (C-4), 30.3 (m, C-3), 47.4 (C-5), 63.0 (C-2) and 174.0 (acid).

(2*S*,3*R*)-[2,3-²H₂]-Proline (1b)

This was prepared as above from methyl (2S,3R)- $[2,3^{-2}H_2]$ -*N*-*tert*-butoxycarbonylprolinate **12b** (42 mg, 0.18 mmol) (22 mg, 94%); mp 230 °C (decomp.); $[a]_D^{26} - 85.2$ (*c* 0.20, H₂O); *m/z* [+ve FAB (thioglycerol)] 140 ([M + Na]⁺) and 118 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 3400 (br, NH and OH) and 1626 (carboxylate); δ_H (360 MHz, C²H₃O²H) 1.95 (2H, m, H-4), 2.20 (1H, br t, H-3*S*), 3.23 (1H, m, H-5A) and 3.36 (1H, m, H-5B); δ_C (62.9 MHz, C²H₃O²H) 24.0 (C-4), 28.9 (t, C-3), 46.0 (C-5) and 172.9 (acid). The signal for C-2 was too weak to detect.

tert-Butyl (2S)-N-tert-butoxycarbonyl-4-oxoprolinate (13)

This was prepared as described in ref. 14; $\delta_{\rm H}$ (360 MHz, C²HCl₃), 1.42 [18H, 2s, C(CH₃)₃], 2.5 (1H, d, $J_{3S,3R}$ 18.8, H-3S), 2.89 (1H, m, $J_{3R,2}$ 10.4, $J_{3R,3S}$ 18.8, H-3*R*), 3.85 (2H, m, H-5) and 4.55 and 4.63 (1H, 2 × br d, $J_{2,3R}$ 10.4, H-2); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃), 27.88 and 28.23 [C(CH₃)₃], 40.87 and 41.40 (C-3), 52.51 and 52.95 (C-5), 56.56 and 57.03 (C-2), 81.03 and 82.36 [OC(CH₃)₃], 153.65 and 154.34 (urethane), 170.84 and 170.92 (ester) and 208.18 and 209.0 (ketone). Irradiation at the two doublets for H-2 at δ 4.55 and 4.63 ppm gave a 4.5% enhancement to the multiplet at δ 2.89 ppm, thus defining this as the signal due to H-3*R*.

tert-Butyl (2*S*,3*R*)-[3-²H₁]-*N*-*tert*-butoxycarbonyl-4-oxoprolinate (13a)

Diisopropylamine (2.9 ml, 20.7 mmol) was dissolved in dry tetrahydrofuran (50 ml) and cooled to -78 °C under nitrogen. n-Butyl lithium (1.6 M in hexane, 11.6 ml, 18.56 mmol) was added and the solution was stirred for 30 min. A solution of tert-butyl (2S)-N-tert-butoxycarbonyl-4-oxoprolinate 13 (5 g, 17.5 mmol) in dry tetrahydrofuran (50 ml) was added and the mixture was stirred for 1 h at -78 °C. Chlorotrimethylsilane (3.8 ml, 297 mmol) was added slowly over a period of 5 min. The mixture was allowed to warm to room temperature, stirred for 1 h and deuterium oxide (15 ml) was added. The organic solvents were removed in vacuo and the aqueous residue was extracted with ethyl acetate $(3 \times 50 \text{ ml})$ and dried (MgSO₄). The solvent was removed *in vacuo* to give *tert*-butyl (2S, 3R)- $[3^{-2}H_{1}]$ -*N*-*tert*-butoxycarbonyl-4-oxoprolinate 13a (5.27 g, quant.) as a pale yellow oil which was reduced without further purification as described below; v_{max} (film)/cm⁻¹ 1769 (ketone), 1742 (ester) and 1708 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.42 [18H, br s, C(CH₃)₃], 2.5 (1H, s, H-3S), 3.8 (2H, m, H-5) and 4.52 and 4.6 (1H, 2s, H-2).

tert-Butyl (2*S*)-[3,3-²H₂]-*N*-*tert*-butoxycarbonyl-4-oxoprolinate (13c)

Deuterium oxide (15 ml) and silica gel prewashed with deuterium oxide (3 \times 10 ml) were added to a solution of *tert*-butyl (2*S*)-*N*-*tert*-butoxycarbonyl-4-oxoprolinate **13** (6 g, 21 mmol) in dry tetrahydrofuran (15 ml). The mixture was stirred at room temperature for 10 days under nitrogen, filtered, and the filtrate

was concentrated *in vacuo*. The aqueous phase was extracted with ethyl acetate (3 × 50 ml) and the combined organic layers were washed with saturated aqueous sodium chloride (50 ml) and dried (MgSO₄). The solvent was removed *in vacuo* to afford *tert*butyl (2*S*)-[3,3-²H₂]-*N*-*tert*-butoxycarbonyl-4-oxoprolinate **13c** as a colourless oil (5.6 g, 93%) which was not further purified; $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.46 and 1.47 [18H, 2s, C(CH₃)₃], 3.86 (2H, m, H-5) and 4.57 (50%) and 4.65 (50%) (1H, 2 br s, H-2); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 27.85 [C(CH₃)₃], 28.19 [C(CH₃)₃], 40.93 (m, C-3), 52.49 and 52.92 (C-5), 56.44 and 56.90 (C-2), 80.97 and 82.31 [OC(CH₃)₃], 153.63 and 154.31 (urethane), 170.82 and 170.91 (ester) and 208.23 and 209.0 (ketone).

tert-Butyl (2S,4S)-N-tert-butoxycarbonyl-4-hydroxyprolinate (15)

tert-Butyl (2S)-N-tert-butoxycarbonyl-4-oxoprolinate 13 (4.8 g, 17 mmol) was dissolved in distilled methanol (51 ml) and dry diethyl ether (66 ml) and the solution was cooled to 0 °C under nitrogen. Sodium borohydride (1.32 g, 35 mmol) in distilled methanol (51 ml) and dry diethyl ether (66 ml) was added and the reaction was stirred at 0 °C for 5 min and allowed to warm to room temperature for 30 min. The solvent was removed in vacuo and the residue was dissolved in ethyl acetate (60 ml) and washed with saturated aqueous sodium hydrogen carbonate (2×60 ml), water (2 \times 60 ml) and saturated aqueous sodium chloride (2 \times 60 ml) and dried (MgSO₄). The solvent was removed in vacuo to give tert-butyl (2S,4S)-N-tert-butoxycarbonyl-4-hydroxyprolinate **15** as a white solid (4.77 g, 98%); mp 53–55 °C; $[a]_{D}^{34}$ –2.52 (c 0.5, CHCl₃); (Found C, 58.6; H, 9.0; N, 4.85. C₁₄H₂₅NO₅ requires C, 58.5; H, 8.8; N, 4.9%); m/z [+ve FAB (3-NBA)] 288 ([M + H]⁺); $v_{\rm max}$ (KBr)/cm⁻¹ 1745 (ester) and 1674 (urethane); $\delta_{\rm H}$ (500 MHz, C²HCl₃) 1.45 [18H, 3s, C(CH₃)₃], 2.0 (1H, 2d, J_{3S,3R} 11.5, H-3S), 2.3 (1H, m, J_{3R4} 9.6, J_{3R2} 9.8, J_{3R35} 11.5, H-3R), 3.47–3.75 (3H, m, 1H exch., J_{5R4} 4.6, J_{5R,5S} 11.9, J_{5S,5R} 11.9, OH and H-5), 4.16 (63%) and 4.22 (37%) (1H, 2d, J_{2,3R} 9.8, H-2) and 4.3 (1H, m, J_{4,5R} 4.6, $J_{4,3R}$ 9.6, H-4); irradiation at δ 4.16 and 4.22 ppm (H-2) caused an 11.1% enhancement at δ 2.3 ppm and a 1.6% enhancement at δ 2.0 ppm, indicating that the former was due to H-3R; irradiation at δ 4.3 ppm (H-4) caused a 9.2% enhancement at δ 2.3 ppm (H-3R) and a 3.3% enhancement at δ 2.0 ppm (H-3S), confirming the stereochemistry at C-4; $\delta_{\rm H}$ (360 MHz, C²H₃CN, 60 °C) 1.45 [18H, 2s, C(CH₃)₃], 1.89 (1H, m, J_{35.5A} 1.3, J_{35.4} 3.5, J_{35.2} 3.4 J_{35.3R} 13.6, H-3S), 2.35 (1H, m, J_{3R4} 5.4, J_{3R2} 9.4 J_{3R3S} 13.6, H-3R), 3.08 (1H, d, exch., J_{OH4} 6.3, OH), 3.28 (1H, ddd, J_{5A,3S} 1.3, J_{5A,4} 3.1, J_{5A,5B} 11.4, H-5A) 3.55 (1H, dd, J_{5B,4} 5.3, J_{5B,5A} 11.4, H-5B), 4.13 (1H, dd, $J_{2,3S}$ 3.5, $J_{2,3R}$ 9.4, H-2) and 4.28 (1H, m, H-4); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 27.80 and 27.87 [C(CH₃)₃], 28.30 and 28.45 [C(CH₃)₃], 37.67 and 38.64 (C-3), 55.67 and 56.08 (C-5), 58.72 and 58.75 (C-2), 70.33 and 71.31 (C-4), 80.08 and 80.24 [OC(CH₃)₃], 82.32 and 82.50 [OC(CH₃)₃], 153.82 and 154.33 (urethane) and 174.3 and 174.53 (ester).

tert-Butyl (2*S*,3*R*,4*S*)-[3⁻²H₁]-*N*-*tert*-butoxycarbonyl-4hydroxyprolinate (15a)

This was prepared from *tert*-butyl (2S,3R)- $[3-^2H_1]$ -*N-tert*-butoxycarbonyl-4-oxoprolinate **13a** (4.8 g, 17 mmol) as described above to afford a white solid (4.17 g, 85%). A small amount was purified for analysis by column chromatography on silica gel, eluting

with ethyl acetate–petroleum ether (1 : 1); mp 55–57 °C; $[a]_{D}^{32}$ –4.38 (*c* 0.5, CHCl₃); *m/z* [+ve FAB (EtOAc/3-NBA)] 289 ([M + H]⁺); ν_{max} (KBr)/cm⁻¹ 1748 (ester) and 1654 (urethane); δ_{H} (360 MHz, C²HCl₃) 1.48 [18H, 3s, C(CH₃)₃], 2.03 (1H, br s, H-3*S*), 3.46–3.79 (3H, m, 1H exch., OH and H-5), 4.19 (1H, 2s, H-2) and 4.3 (1H, br d, H-4); δ_{C} (125.8 MHz, C²HCl₃) 27.79 and 27.85 [C(CH₃)₃], 28.29 [C(CH₃)₃], 38.3 (m, C-3), 55.61 and 56.02 (C-5), 58.67 and 58.3 (C-2), 70.21 and 71.2 (C-4), 80.05 and 80.22 [OC(CH₃)₃], 82.27 and 82.45 [OC(CH₃)₃], 153.82 and 154.31 (urethane) and 174.22 and 174.45 (ester).

tert-Butyl (2*S*,3*S*,4*S*)-[3-²H₁]-*N*-*tert*-butoxycarbonyl-4-hydroxyprolinate (15b)

Diisopropylamine (2.19 ml, 16 mmol) was dissolved in dry tetrahydrofuran (38 ml) and cooled to -78 °C under nitrogen. *n*-Butyl lithium (1.6 M in hexane; 8.76 ml, 14 mmol) was added and the solution was stirred for 30 min. A solution of tert-butyl (2S)- $[3,3^{-2}H_2]$ -*N-tert*-butoxycarbonyl-4-oxoprolinate **13c** (3.9 g, 14 mmol) in dry tetrahydrofuran (38 ml) was added and the mixture was stirred for 1 h at -78 °C. Chlorotrimethylsilane (2.87 ml, 22 mmol) was added slowly over a period of 5 min. The reaction was allowed to warm to room temperature, stirred for 1 h and poured into saturated aqueous ammonium chloride (50 ml). The solvent was concentrated in vacuo. The aqueous layer was extracted with ethyl acetate (3 \times 50 ml) and dried $(MgSO_4)$. The solvent was removed in vacuo to produce (2S,3S)-[3-²H₁]-*N*-tert-butoxycarbonyl-4-oxoprolinate **13b** (3.9 g, quant.) as a pale yellow oil which was reduced without purification using the method described above. The product was purified by column chromatography on silica gel, eluting with ethyl acetatepetroleum ether (1 : 1) to give tert-butyl (2S,3S,4S)- $[3-^{2}H_{1}]$ -Ntert-butoxycarbonyl-4-hydroxyprolinate 15b as a colourless oil $(1.59 \text{ g}, 40\%); [a]_{D}^{32} - 6.32 (c \ 0.5, \text{CHCl}_3); m/z \text{ [ES+] } 327 ([M +$ K]⁺), 311 ([M + Na]⁺) and 289 ([M + H]⁺); v_{max} (film)/cm⁻¹ 1745 (ester) and 1674 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.46 [18H, 3s, C(CH₃)₃], 2.28 (1H, m, H-3R), 3.47–3.75 (3H, m, 1H exch., OH and H-5), 4.1–4.3 (H-2 and H-4); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 27.92 [C(CH₃)₃], 28.34 [C(CH₃)₃], 38.2 (m, C-3), 55.79 and 56.9 (C-5), 58.75 (C-2), 70.38 and 71.37 (C-4), 80.3 and 80.29 [OC(CH₃)₃], 82.39 and 82.58 [OC(CH₃)₃], 153.87 (urethane) and 174.36 (ester).

tert-Butyl (2*S*,4*S*)-[3,3-²H₂]-*N*-*tert*-butoxycarbonyl-4hydroxyprolinate (15c)

tert-Butyl (2*S*)-[3,3-²H₂]-*N*-*tert*-butoxycarbonyl-4-oxoprolinate **13c** (2.55 g, 8.9 mmol) was reduced according to the method described above to yield *tert*-butyl (2*S*,4*S*)-[3,3-²H₂]-*N*-*tert*-butoxycarbonyl-4-hydroxyprolinate **15c** as a colourless oil (2.28 g, 89%), *m*/*z* [+ve FAB (3-NBA)] 290 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 3430 (OH), 1745 (ester) and 1703 (br, urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.5 [18H, 3s, C(CH₃)₃], 3.49–3.77 (3H, m, 1H exch., OH and H-5), 4.21 (1H, 2s, H-2) and 4.3 (1H, m, H-4); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 27.90 and 28.3 [C(CH₃)₃], 38.4 (m, C-3), 55.67 and 56.08 (C-5), 58.72 and 58.75 (C-2), 70.33 and 71.31 (C-4), 80.08 and 80.24 [OC(CH₃)₃], 82.32 and 82.50 [OC(CH₃)₃], 153.82 and 154.33 (urethane) and 174.3 and 174.53 (ester).

tert-Butyl (2*S*,4*S*)-*N*-*tert*-butoxycarbonyl-4-*O*-*para*-toluenesulfonyloxyprolinate (17)

tert-Butyl (2S,4S)-N-tert-butoxycarbonyl-4-hydroxyprolinate 15 (4.5 g, 15.7 mmol) was dissolved in pyridine (21 ml, 260 mmol) and cooled to -15 °C under nitrogen. para-Toluenesulfonyl chloride (10.11 g, 53 mmol) was added over 30 min and the mixture was stirred at room temperature for 20 h. The solvent was removed in vacuo and the resultant solid was partitioned between ethyl acetate (60 ml) and water (60 ml). The organic phase was washed with 10% aqueous citric acid (60 ml), water (60 ml) and saturated aqueous sodium chloride (30 ml) and dried (MgSO₄). The solvent was removed in vacuo and the residue was purified by column chromatography on silica gel, eluting with ethyl acetate-petroleum ether (1:2) to afford tert-butyl (2S,4S)-N-tert-butoxycarbonyl-4-O-para-toluenesulfonyloxyprolinate 17 as a colourless oil which solidified on standing to give a white solid (3.98 g, 58%); mp 67– 70 °C; [*a*]_D³⁴ –28.34 (*c* 0.5, CHCl₃); (Found C, 57.0; H, 7.2; N, 3.1. C₂₁H₃₁NO₇S requires C, 57.1; H, 7.1; N, 3.2%); *m/z* [+ve FAB (ethyl acetate/3-NBA)] 442 ($[M + H]^+$); v_{max} (KBr)/cm⁻¹ 1707 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.5 [18H, 4s, C(CH₃)₃], 2.4 (2H, m, H-3), 2.45 (3H, s, CH₃), 3.5 (1H, t, J_{5A.5B} 13.0, H-5A), 3.65 (1H, m, J_{5B,4} 5.5, J_{5B,5A} 13.0, H-5A), 4.2 (54%) and 4.3 (46%) (1H, 2 dd, $J_{\scriptscriptstyle 2,3S}$ 2.8, $J_{\scriptscriptstyle 2,3R}$ 9.07, H-2), 5.0 (1H, m, $J_{\scriptscriptstyle 4,5R}$ 5.5, H-4), 7.35 (2H, 2d, J_{ortho} 8.1, ArH) and 7.76 (1H, 2 d, J_{ortho} 8.1, ArH); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 21.04 and 21.65 (CH₃), 27.79, 28.25 and 28.30 [C(CH₃)₃], 36.03 and 37.18 (C-3), 51.81 and 52.25 (C-5), 57.93 and 58.18 (C-2), 78.95 (C-4), 80.21 and 80.31 [OC(CH₃)₃], 81.69 and 81.74 [OC(CH₃)₃], 127.74–145.08 (Ar), 153.42 and 153.59 (urethane) and 170.09 and 170.40 (ester).

tert-Butyl (2*S*,3*R*,4*S*)-[3-²H₁]-*N*-*tert*-butoxycarbonyl-4-*O*-*para*-toluenesulfonyloxyprolinate (17a)

This was prepared from *tert*-butyl (2*S*,3*R*,4*S*)-[3⁻²H₁]-*N*-*tert*-butoxycarbonyl-4-hydroxyprolinate **15a** (3.96 g, 14 mmol) as described above (2.04 g, 33%); mp 69–71 °C; $[a]_D^{32}$ –23.68 (*c* 0.5, CHCl₃); (*m*/*z* [EI] Found 442.1911 C₂₁H₃₀²HNO₇S requires 442.1884); *v*_{max} (KBr)/cm⁻¹ 1742 (ester) and 1706 (urethane); δ_H (360 MHz, C²HCl₃) 1.45 [18H, 4s, C(CH₃)₃], 2.36 (1H, m, H-3*S*), 2.45 (3H, s, CH₃), 3.6 (2H, m, H-5), 4.2 (54%) and 4.3 (46%) (1H, 2s, H-2), 5.0 (1H, br s, H-4), 7.35 (2H, d, *J*_{ortho} 7.5, ArH) and 7.76 (1H, d, *J*_{ortho} 7.5, ArH); δ_C (125.8 MHz, C²HCl₃) 21.64 (CH₃), 27.81, 28.26 and 28.31 [C(CH₃)₃], 37.18 (m, C-3), 51.81 and 52.25 (C-5), 57.89 and 58.14 (C-2), 77.93 and 78.74 (C-4), 80.22 and 80.32 [OC(CH₃)₃], 81.74 [OC(CH₃)₃], 127.75–145.07 (Ar), 153.44 and 153.61 (urethane) and 170.11 and 170.41 (ester).

tert-Butyl (2*S*,3*S*,4*S*)-[3-²H₁]-*N*-*tert*-butoxycarbonyl-4-*O*-*para*-toluenesulfonyloxyprolinate (17b)

This was prepared from *tert*-butyl (2*S*,3*S*,4*S*)-[3-²H₁]-*N*-*tert*butoxycarbonyl-4-hydroxyprolinate **15b** (1.5 g, 5.2 mmol) as described above to give the product as a colourless oil (1.64 g, 71%); $[a]_D^{34} - 21.48$ (*c* 0.25, CHCl₃); *m*/*z* [EI] 442 ([M]⁺) v_{max} (KBr)/cm⁻¹ 1700 (br, urethane); δ_H (360 MHz, C²HCl₃) 1.5 [18H, 4s, C(CH₃)₃], 2.45 (4H, s + m, CH₃ and H-3*R*), 3.53 (1H, m, *J*_{5A,4} 2.25 *J*_{5A,5B} 13.0, H-5A), 3.59 (1H, m, *J*_{5B,4} 5.64, *J*_{5B,5A} 13.0, H-5B), 4.21 (67%) and 4.33 (33%) (1H, 2d, *J*_{2,3R} 9.06, H-2), 5.0 (1H, br s, H-4), 7.35 (2H, 2d, *J*_{ortho} 8.0, ArH) and 7.8 (1H, 2d, *J*_{ortho} 8.0, ArH); δ_C (125.8 MHz, $C^{2}HCl_{3}$ 21.64 (CH₃), 27.81, 28.26 and 28.31 [C(CH₃)₃], 37.18 (m, C-3), 51.81 and 52.25 (C-5), 57.95 and 58.14 (C-2), 77.93 and 78.88 (C-4), 80.22 and 80.32 [OC(CH₃)₃], 81.69 and 81.74 [OC(CH₃)₃], 127.75–145.07 (Ar), 153.43 and 153.61 (urethane) and 170.1 and 170.4 (ester).

tert-Butyl (2S,4S)-[3,3- $^{2}H_{2}$]-N-tert-butoxycarbonyl-4-O-paratoluenesulfonyloxyprolinate (17c)

This was prepared from *tert*-butyl (2S,4S)- $[3,3^{-2}H_2]$ -*N-tert*butoxycarbonyl-4-hydroxyprolinate **15c** (2.28 g, 7.9 mmol) as described above (1.75 g, 50%); mp 67–70 °C; *m/z* [ES+] 444 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 1747 (urethane); $\delta_{\rm H}$ (360 MHz, C²HCl₃) 1.5 [18H, 4s, C(CH₃)₃], 2.45 (3H, s, CH₃), 3.6 (2H, m, H-5), 4.25 (1H, 2s, H-2), 5.0 (1H, br s, H-4), 7.35 (2H, 2d, *J*_{ortho} 7.5, ArH) and 7.76 (1H, 2d, *J*_{ortho} 7.5, ArH); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 21.64 (CH₃), 27.80 [C(CH₃)₃], 28.25 [C(CH₃)₃], 36.0 (C-3), 51.81 and 52.25 (C-5), 57.81 and 58.06 (C-2), 77.89 and 78.83 (C-4), 80.26 and 80.35 [OC(CH₃)₃], 81.77 [OC(CH₃)₃], 127.75–145.09 (Ar), 153.46 (urethane) and 170.43 (ester).

tert-Butyl (2S)-N-tert-butoxycarbonylprolinate (18)

tert-Butyl (2S,4S)-N-tert-butoxycarbonyl-4-O-para-toluenesulfonyloxyprolinate 17 (3.55 g, 8.05 mmol) was dissolved in dry dimethyl sulfoxide (60 ml). Sodium borohydride (1.87 g, 49 mmol) was added and the mixture was heated under nitrogen at 85 °C for 6.5 h. The mixture was cooled, diluted with water (100 ml) (effervescence) and the aqueous phase was extracted with diethyl ether (3 \times 100 ml). The combined organic phases were washed with saturated aqueous sodium chloride (50 ml) and dried (MgSO₄). The solvent was removed in vacuo to afford tert-butyl (2S)-N-tert-butoxycarbonylprolinate 18 as a colourless oil (2.04 g, 94%) which was used without further purification. A small amount was purified for analysis by column chromatography on silica gel, eluting with ethyl acetate-petroleum ether (1 : 4); $[a]_{p}^{22}$ -50.2 (c 1, CHCl₃); (m/z [EI] Found 271.17718, C₁₄H₂₅NO₄ requires 271.17838); m/z [ES+] 272 ([M + H]⁺); v_{max} (film)/cm⁻¹ 1742 (ester) and 1703 (urethane); $\delta_{\rm H}$ (250 MHz, C²HCl₃) 1.45 [18H, 2s, C(CH₃)₃], 1.85 (3H, m, H-3R and H-4), 2.19 (1H, m, H-3S), 3.45 (2H, m, H-5) and 4.15 (1H, m, H-2); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 23.37 and 24.16 (C-4), 27.96 and 28.32 [C(CH₃)₃], 28.38 and 29.84 [C(CH₃)₃], 30.84 (C-3), 46.26 and 46.47 (C-5), 59.66 (C-2), 79.38 and 79.58 [OC(CH₃)₃], 80.78 [OC(CH₃)₃], 153.95 and 154.28 (urethane) and 172.29 (ester).

tert-Butyl (2S,3S)-[3-²H₁]-N-tert-butoxycarbonylprolinate (18a)

This was prepared from *tert*-butyl (2*S*,3*R*,4*S*)-[3-²H₁]-*N*-*tert*butoxycarbonyl-4-*O*-*para*-toluenesulfonyloxyprolinate **17a** (2 g, 4.52 mmol) as described above (1.07 g, 87%); $[a]_D^{33}$ –44.94 (*c* 0.95, CHCl₃); *m*/*z* [ES+] 273 ([M + H]⁺); v_{max} (film)/cm⁻¹ 1739 (ester) and 1703 (urethane); δ_H (250 MHz, C²HCl₃) 1.45 [18H, 2s, C(CH₃)₃], 1.85 (3H, m, H-3*R* and H-4), 3.45 (2H, m, H-5) and 4.15 (1H, 2s, H-2).

tert-Butyl (2*S*,3*R*)-[3-²H₁]-*N*-*tert*-butoxycarbonylprolinate (18b)

This was prepared from *tert*-butyl (2S,3S,4S)- $[3-^{2}H_{1}]$ -*N*-*tert*-butoxycarbonyl-4-*O*-para-toluenesulfonyloxyprolinate **17b** (1.6 g,

3.6 mmol) as described above (580 mg, 59%); $[a]_{D}^{33} - 47.87$ (*c* 1, CHCl₃); *m/z* [EI] 272 ([M]⁺); v_{max} (film)/cm⁻¹ 1742 (ester) and 1703 (urethane); δ_{H} (250 MHz, C²HCl₃) 1.45 [18H, 2s, C(CH₃)₃], 1.85 (2H, m, H-4), 2.19 (1H, m, H-3S), 3.45 (2H, m, H-5) and 4.15 (1H, 2d, H-2); δ_{C} (125.8 MHz, C²HCl₃) 23.37 and 24.16 (C-4), 27.96 and 28.32 [C(CH₃)₃], 28.38 and 29.84 [C(CH₃)₃], 30.72 (m, C-3), 46.26 and 46.47 (C-5), 59.66 (C-2), 79.38 and 79.58 [OC(CH₃)₃], 80.78 [OC(CH₃)₃], 153.95 and 154.28 (urethane) and 172.29 (ester).

tert-Butyl (2S)-[3,3-2H2]-N-tert-butoxycarbonylprolinate (18c)

This was prepared from *tert*-butyl (2*S*,4*S*)-[3,3⁻²H₂]-*N*-*tert*-butoxycarbonyl-4-*O*-*para*-toluenesulfonyloxyprolinate **17c** (1.75 g, 3.95 mmol) as described above (960 mg, 89%), m/z [EI] 273 [(M)⁺]; v_{max} (film)/cm⁻¹ 1740 (ester) and 1702 (urethane); δ_{H} (250 MHz, C²HCl₃) 1.45 [18H, 2s, C(CH₃)₃], 1.85 (2H, m, H-4), 3.45 (2H, m, H-5) and 4.15 (1H, 2s, H-2).

(2S)-Proline (1)

tert-Butyl (2S)-N-tert-butoxycarbonylprolinate 18 (720 mg, 2.66 mmol) was stirred with 6 M aqueous hydrochloric acid (12 ml) at room temperature for 2 h. The solvent was removed in vacuo and the residue was dissolved in water (5 ml). The pH was adjusted from pH 2 to 6 using 2 M aqueous ammonium hydroxide and the solution was loaded on to a Dowex 50X8 (H⁺) ion exchange column. The column was eluted with water until the eluant was no longer acidic and the product was eluted with 2 M aqueous ammonium hydroxide. The solvent was removed in vacuo. The residue was dissolved in water (5 ml) and lyophilised to afford (2S)-proline 1 as an off-white solid which was recrystallised from diethyl ether and ethanol (300 mg, 88%); mp 223 °C (decomp.); (lit.¹⁵ 227–229 °C); $[a]_{D}^{23}$ –61.48 (c 1.33, MeOH) [lit.¹⁵ –83 (c 1, H₂O)]; m/z [ES+] 116 ([M + H]⁺); v_{max} (KBr)/cm⁻¹ 1624 (acid); $\delta_{\rm H}$ (500 MHz, C²H₃O²H) 1.96 (2H, m, H-4), 2.1 (1H, m, H-3R), 2.29 (1H, m, H-3S), 3.25 (1H, m, H-5A), 3.38 (1H, m, H-5B) and 4.0 (1H, dd, $J_{2,3R}$ 6.23, $J_{2,3S}$ 8.74, H-2); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 25.63 (C-4), 30.91 (C-3), 47.53 (C-5), 63.16 (C-2) and 174.55 (acid).

(2*S*,3*S*)-[3-²H₁]-Proline (1a)

This was prepared from *tert*-butyl (2*S*,3*S*)-[3-²H₁]-*N*-*tert*butoxycarbonylprolinate **18a** (1 g, 3.68 mmol) as described above and was purified by dissolving in methanol and boiling with decolourising charcoal (126 mg, 27%), mp 225 °C (decomp.); $[a]_{D}^{28}$ -47.2 (*c* 0.25, MeOH); *m/z* [ES+] 117 ([M + H]⁺); *v*_{max} (KBr)/cm⁻¹ 1624 (acid); δ_{H} (360 MHz, C²H₃O²H) 1.96 (2H, m, H-4), 2.1 (1H, m, H-3*R*), 3.25 (1H, m, H-5A), 3.38 (1H, m, H-5B) and 4.0 (1H, d, *J*_{2,3*R*} 5.8, H-2); δ_{C} (125.8 MHz, C²HCl₃) 25.53 (C-4), 30.8 (m, C-3), 48.99 (C-5), 63.09 (C-2) and 174.54 (acid).

(2*S*,3*R*)-[3-²H₁]-Proline (1c)

This was prepared from *tert*-butyl (2S,3R)- $[3^{-2}H_1]$ -*N-tert*butoxycarbonylprolinate **18b** (580 mg, 2.13 mmol) as described above (195 mg, 71%); mp 225 °C (decomp.); $[a]_D^{32}$ –57.36 (*c* 1.1, MeOH); *m/z* [+ve FAB (thioglycerol)] 139 ([M + Na]⁺ and 117 ([M + H]⁺); v_{max} (film)/cm⁻¹ 1626 (acid); δ_H (360 MHz, C²H₃O²H) 1.96 (2H, m, H-4), 2.29 (1H, m, H-3*S*), 3.25 (1H, m, H-5A), 3.38 (1H, m, H-5B) and 4.0 (1H, d, $J_{2,35}$ 8.48, H-2); $\delta_{\rm C}$ (125.8 MHz, C²HCl₃) 25.53 (C-4), 30.8 (m, C-3), 47.52 (C-5), 63.10 (C-2) and 174.47 (acid).

(2S)-[3,3-²H₂]-Proline (1d)

This was prepared from *tert*-butyl (2*S*)-[3,3-²H₂]-*N*-*tert*butoxycarbonylprolinate **18c** (960 mg, 3.51 mmol) as described above and the product was purified by dissolving in methanol and boiling with decolourising charcoal (240 mg, 53%); mp 227 °C (decomp.); $[a]_{D}^{33}$ –63.53 (*c* 1, MeOH) *m*/*z* [+ve FAB (thioglycerol)] 118 ([M + H]⁺); v_{max} (film)/cm⁻¹ 1624 (acid); δ_{H} (360 MHz, C²H₃O²H) 1.96 (2H, m, H-4), 3.25 (1H, m, H-5A), 3.38 (1H, m, H-5B) and 4.0 (1H, s, H-2); δ_{C} (125.8 MHz, C²HCl₃) 25.45 (C-4), 30.6 (m, C-3), 47.54 (C-5), 63.05 (C-2) and 174.55 (acid).

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References

- This work has been published as separate communications, in: (a) P. Dieterich and D. W. Young, *Tetrahedron Lett.*, 1993, 34, 5455–5458;
 (b) P. Barraclough, C. A. Spray and D. W. Young, *Tetrahedron Lett.*, 2005, 46, 4653–4655.
- 2 D. W. Young, Topics Stereochem., 1994, 21, 381-465.
- 3 M. Kainosho, Nat. Struct. Biol., NMR Suppl., 1997, 858-861.
- 4 K. Wuthrich, *NMR of Proteins and Nucleic Acids*, Wiley, New York, 1986.
- 5 G. C. K. Roberts, NMR of Macromolecules. A Practical Approach, Oxford University Press, Oxford, 1993.
- 6 M. Oba, T. Terauchi, J. Hashimoto, T. Tanaka and K. Nishiyama, *Tetrahedron Lett.*, 1997, **38**, 5515–5518; M. Oba, T. Terauchi, A. Miyakawa and K. Nishiyama, *Tetrahedron: Asymmetry*, 1999, **10**, 937–945; M. Oba, A. Miyakawa, K. Nishiyama, T. Terauchi and M. Kainosho, J. Org. Chem., 1999, **64**, 9275–9278.
- 7 (a) S. J. Field and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1983, 2387–2392; (b) D. Gani and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1983, 2393–2398; (c) D. Gani, D. W. Young, D. M. Carr, J. P. Poyser and I. H. Sadler, J. Chem. Soc., Perkin Trans. 1, 1983, 2811–2814.
- 8 D. Gani and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1985, 1355– 1362; D. Gani, P. B. Hitchcock and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1985, 1363–1372.
- 9 D. W. Young, in *Isotopes in the Physical and Biomedical Sciences, vol. 1, Labelled Compounds (Part B)*, E. Buncel and J. R. Jones, eds., Elsevier, Amsterdam, 1991, pp. 341–427.
- 10 K. M. Lee, K. Ramalingam, J.-K. Son and R. W. Woodard, J. Org. Chem., 1989, 54, 3195–3198.
- B. S. Axelsson, K. J. O'Toole, P. A. Spencer and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1994, 807–816; B. S. Axelsson, N. J. Church, P. A. Spencer and D. W. Young, Folia Microbiol. (Prague, Czech Repub.), 1995, 40, 17–22; K. J. M. Beresford and D. W. Young, Tetrahedron, 1996, 52, 9891–9900; N. J. Church and D. W. Young, J. Chem. Soc., Perkin Trans. 1, 1998, 1475–1482.
- 12 F. Weygand, H. J. Bestmann and E. Klieger, *Chem. Ber.*, 1958, **91**, 1037–1040; H. A. DeWald and A. M. Moore, *J. Am. Chem. Soc.*, 1958, **80**, 3941–3945.
- 13 J. Hondrelis, G. Lonergan, S. Voliotis and J. Matsoukas, *Tetrahedron*, 1990, 46, 565–576.
- 14 P. Barraclough, P. Hudhomme, C. A. Spray and D. W. Young, *Tetrahedron*, 1995, **51**, 4195–4212.
- 15 K. Drauz, A. Kleeman, J. Martens, P. Scherberich and F. Effenberger, J. Org. Chem., 1986, 51, 3494–3498.
- 16 T. Katoh, Y. Nagata, Y. Kobayashi, K. Arai, J. Minami and S. Terashima, *Tetrahedron*, 1994, 50, 6221–6238.
- 17 J.-R. Dormoy and B. Castro, Synthesis, 1986, 81-82.